

&KDUDFWHULJDWLRQRI F\VWDWKLRQLQH O\DVH IURP T.gondii \$ W

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Toxoplasma gondii is a protozoan parasite of medical and veterinary relevance responsible for toxoplasmosis in humans. As there is currently no vaccine available for human, the identification of good target candidates for future drug development is urgently required. A recent proteomic analysis of partially sporulated oocysts of *T. gondii* showed that oocysts have a greater capability of de novo amino acid biosynthesis, shedding light on a stage-specific subset of proteins whose functional profile is consistent with the oocyst need to resist various environmental stresses. Among these putative oocyst/sporozoite-specific proteins, three enzymes involved in cysteine metabolism, i.e., cystathionine γ -synthase, cystathionine γ -lyase (CGL) and cysteine synthase, were found. However, despite the central metabolic roles of these enzymes, the functionality of none of them has so far been investigated. Herein, CGL from *T. gondii* (TgCGL) has been cloned, expressed and physicochemically and enzymatically characterized. The purified TgCGL is a functional enzyme which splits L-cystathionine almost exclusively at the C-S bond to yield L-cysteine. This finding likely implies that the reverse transsulfuration pathway is operative in the parasite. The enzyme displays only marginal reactivity toward L-cysteine, which is also a mixed-type inhibitor of TgCGL activity, therefore indicating a tight regulation of cysteine intracellular levels in the parasite. Structure-guided homology modelling revealed two striking amino acid differences between human and parasite CGL active sites.

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