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*ostreatus*, A-6 and OST-1 showed higher reduction of 84% and 64% respectively. *Pleurotus* spp. (Figure 2) showed isolates OST-1 was most effective in HCH persistence respectively. Exotic strains of same species used produced 67% and 65% quantitative reduction in 4,4-DDT and were OST-Belgium and WC-522 which manifested less reduction of 21% and 24% respectively. In presence of A-6 (69%), OST-Belgium (69%) 29% and 45% respectively (Table 3). Least reduction (4%) was observed in case of WC-537 (51%) was more pronounced for 4,4-DDT than for 2,4-DDT by using *Pleurotus Pulmonarius* (WC-537) compared with exotic and local isolates of *Pleurotus ostreatus*. The order of biodegradation of HCH in presence of white rot fungi species were OST-1>A6>WC-522>OST-Belgium>WC-537. Likewise, -HCH was highly degraded by A-6(85%) followed by OST-1(45%) while minimum reduction was observed when OST-Belgium was used in bioassay suggesting transformation of HCH cannot be degraded by this specific isolate of *Pleurotus ostreatus*. -HCH was degraded in following manner OST-1>A-6>WC-522>WC-537>OST-Belgium.

Biodegradation rate of different isomer of OCPs depends on microorganism spp. and aeration. Likewise HCH isomers vary greatly in their persistence and other properties including (solubility, volatility) so they are degraded at different rates and with different ratios even using similar white rot fungi. Inter conversion of isomers into metabolites is also a prime cause of difference in reduction quantities of these HCH isomers by individual isolates. Conversion of HCH isomers to other breakdown products due to biotic factors is also another cause. Conversion of HCH into tetrachlorocyclohexane (TCCOL) and TCCH with an allelic hydroxyl group due to white rot fungi is also reported [22]. Degradation of HCH to TCCOL is attributed to certain active enzymes [8]. Difference in biodegradation efficacy of *Pleurotus ostreatus* has been linked with Lignolytic enzymes production ability. Different enzymes isolated from *Pleurotus ostreatus* could participate in degradation of Lindane and its metabolite in soil [23]. As a genetic character, these enzymes could vary among different strains and species of *Pleurotus* and therefore, can be attributed to the variation in bioremediation potential. Genetic Diversity of the Edible Mushroom *Pleurotus* sp. By using Amplified Fragment Length Polymorphism (AFLP) markers revealed that even geographically close *P. ostreatus* strain from culture collections are genomically distant (their AFLP profile similarity is only 39%), suggesting local and exotic strains also exhibit difference in genetics [24]. Genomic distinction can be another prime cause.

Isolates of *P* spp. were also studied for their potential to degrade 4,4-DDT and 2,4-DDT. Five isolates used in the study were found to have significantly different effects on 4,4-DDT and 2,4-DDT isomers. Mean reduction in 4,4-DDT and 2,4-DDT isomers by different isolates



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OST Belgium (69%) and WC-537 (51%) was more pronounced for 4,4-DDT than for 2,4-DDT with only 14%, 16% and 0% respectively. Exotic strain of *P. ostreatus* (WC-522) was the most inefficient of the isolates and produced only 3% reduction in 2,4-DDT and no reduction was

observed in case of 4,4-DDT. Efficacy of isolates varied significantly with each other for *γ*-endosulfan and *δ*-endosulfan. A-6 and OST-1 were found most efficient and at par with each other in reducing the amount of *γ*- and *δ*-endosulfan in bioassay. Mean maximum reduction of Endrin 32% was recorded in WC-537 followed by 29% in WC-522 and 14% in OST-Belgium.

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