

Keywords Candida parapsilosis complex; Candida parapsilosis sensu stricto; Oral dysbiosis; Immunological status

Introduction

Results on the distribution of the species in this complex are highly variable, although all the literature we reviewed reports *Candida parapsilosis* sensu stricto as having the highest prevalence worldwide, and Silva et al. [1] report it as the most frequent isolate in hematogenous infections. The distribution of *Candida orthopsilosis* and *Candida metapsilosis* varies widely according to geographic region, clinical service and anatomical site [2,3]. Indeed, Miranda et al. [4] claim that the exact importance of *C. orthopsilosis* and *C. metapsilosis* as human pathogens is as yet uncertain.

Little is known regarding the prevalence of species of the *Candida parapsilosis* complex in the oral cavity. The few papers published on the subject report variable results on its distribution in this specific ecological niche, and state that *C. parapsilosis* sensu stricto is the most commonly

outpatients and hospitalized immunocompromised patients in different clinical situations.

The sample consisted of 101 isolates which were successfully reconstituted, to be analyzed by end-point PCR with specific primers. The following reference strains from the ATCC collection were used: *C. parapsilosis* (ATCC 22019), *C. orthopsilosis* (ATCC 96139) and *C. metapsilosis* (ATCC 96143), on which the same procedures were performed as on the clinical isolates.

For clinical correlation, patient's clinical records and the dental data of the oral isolates were available.

For their vitrosusceptibility tests, we used Vitek2 automated susceptibility testing cards AST-YS07 to evaluate the response of 50 clinical isolates to the following antifungal agents: fluconazole (FLC), voriconazole (VRC), caspofungin (CASPO), micafungin (MICA) and amphotericin B (AMB). To interpret the readings, we used the 2012 revision of species-specific clinical breakpoints (CBPs) and epidemiological cut-off value (ECV) (Pfaller and Diekema 2012) [9-12] (CLSI, M27-S4/2012). For quality controls for the study we used the following *Candida* strains: *Candida krusei* (ATCC 6258), *C. parapsilosis* ATCC 22019 and *C. albicans* ATCC 9002.

The following variables were analyzed: a) species of the parapsilosis complex; b) oral ecological niche; c) oral clinical status; d) intraoral appliances; e) immunological status; and f) response to antifungal agents.

Reconstitution of clinical isolates

Isolates were initially identified based on the color developed in the chromogenic medium, micro-morphology in 1%-Tween 80 milk agar and carbohydrate assimilation profile using commercial systems API ID 32D and Vitek2 (BioMérieux, France) [7,9].

To reconstitute the isolates, each strain was seeded in the following culture media: 1) BHI (brain-heart infusion) for metabolic activation of strains, incubated at 28°C-37°C for 24-48 h [7]; 2) Differential chromogenic solid medium for *Candida* (Chromagar), to ascertain the purity of the isolate and discard any contaminated strains, incubated at 28°C for 24 hours [10]; 3) Sabouraud, to amplify colonies, incubated at 28°C-37°C for 24 hours [10]; 4) YPD broth (yeast extract, peptone and glucose) to obtain a more robust culture, for 24 h with shaking at 37°C [10].

Molecular characterization of clinical isolates by end-point PCR with specific primers

Yeast DNA was obtained by breaking down the cell wall with zMd.6(do)16(.8 37°C)TJ2Te(a)-5(k)j5(g)8(en)1(er)te aphorcup7.1(l)-3(a)1(s)5(t (CS-14(a)6(h)4(e;)13(e)ra)9(n)4(d dS14(b) PS14(b)7(h)4(oe)13(o)11(tp7(l)-3(a)1(s)5(t)8(er)13(e)v8(er)16(i)e)-5(d))16(y)tp71(t)-5(i)al micro-ctp71(y 73., f)9(n 6(Pp12(r)13(o)11(t)-6ocol. e ia btained s reteted at

T0le -5(c)-7(tu-5(l)-2.9(a)8(9(r t)-6(y p12(in)1(g)a)3(s)d)12(n)4(e))16(y en)4(d-p)-9.9(o)12(nn)19(t PCR w)-3(1(in1

For phylogenetic analysis we used the BIOEDIT software for editing sequence alignment and the MEGA 6 software for multiple alignment of sequences and phylogenetic analysis, for which the Neighbour joining algorithm was used. The tree was constructed with the reference ATCC sequences for *C. parapsilosis*, *C. orthopsilosis* and *C. metapsilosis* in addition to the sequences selected at random from the total which were positive to PCR with specific primers.

Results

Of the total isolates upon which molecular analysis was performed, 96 (95%) were positive for the *Candida parapsilosis sensu stricto* (Table 2) according to the end-point PCR method with the pair of specific primers CPAR-CPAF, providing 379 bp amplicons (Figures 2-5). This band pattern is compatible to the one published by Asadzadeh et al. in the Journal of Medical Microbiology in 2009 [10]. The 5 remaining strains were negative for all three species of the parapsilosis complex, so they could only be

ere is almost 4 times more probability of recovering Candida parapsilosis of buccal cavity in pathological conditions than in health condition. The difference was statistically significant and clinically relevant (Table 4).

The probability of recovering Candida parapsilosis from the oral cavity with

- Mouth and skin may be reservoirs for *Candida parapsilosis* strains with resistant phenotype and/or reduced susceptibility to the group of azoles, echinocandins and cytosine.

Recommendations

Depending on the limitations of this study, we suggest:

- Repeating this study design but in a prospective model and with a larger sample size to minimize the random error inherent to the process.
- Validate the results of in vitro antifungal sensitivity with the reference method (CLSI M27-S4 / 2008)
- Evaluate sensitivity and specificity of the molecular technique employed in this study to discriminate at species level, by comparing it to the Gold standard in a large number of samples.
- Study the impact of the oral microenvironment in dysbiosis on the virulence of *Candida parapsilosis sensu stricto*.

References

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