

Eco-Friendly Nano-emulsion Formulation of *Mentha piperita* Against Stored Product Pest *Sitophilus oryzae*

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the of *M. piperita* (EO nano-emulsion) as possible as protecting agents of wheat grains against infestation by the rice weevil, *Sitophilus oryzae*

Materials and Methods

Insect rearing

Cultures of the rice weevil, *Sitophilus oryzae* (L.), was maintained in (Stored Products Department, Plant Protection Research Institute, Agriculture Research Center, Dokki, Giza, Egypt.) over 5 years without exposure to insecticides and reared on sterilized whole wheat. Insect rearing and all experimental procedures were carried out at 26.1°C and 65.5% R.H. Adults used in studies was two weeks post-eclosion.

Materials

Tween 80 Acetone and Peppermint oil was purchased as pure oil from Department of Botany and microbiology, Faculty of Science, Alexandria University, Egypt.

Preparation of nano-emulsion

Preparation of nano-emulsion was conducted in National Research Center laboratory, the oil-in-water nano-emulsion was formulated using *M. piperita* (peppermint) essential oil, non-ionic surfactant (tween 80) and deionized water; according to Ghotbi et al. [19] and Sugumar et al. [20]. concentration of *M. piperita* (EO) (4%, v/v) was prepared. Initially, coarse emulsion was prepared by adding water to organic phase containing oil and surfactant in ratios 1:5 (v/v) using a magnetic stirrer; which was then subjected to ultrasonic using a 20kHz Sonicator (*BANDELIN Sonopuls*).

M. piperita nano-emulsion characterizations

Droplet size determined at central lab in National Research Center:

emulsion droplet size and size distribution was determined using particle size analyzer (Malvern-UK, 4700 model) Droplet size was analyzed using dynamic light scattering (DLS) technique [21]. Prior to all the experiments, the nano-emulsion oil formulations were diluted with water to get rid of the multiple scattering droplet size and the polydispersity index (PDI) of the formulated nano-emulsion oil were measured.

Morphology of *M. piperita* nano-emulsion: To visualize the shape and morphology of the formulated *M. piperita* nano-emulsion oil, transmission electron microscopy (TEM) at the EM Unit in the Faculty of Science, Alex. Univ. was carried out. One drop of emulsion was negatively stained with ethanol and was positioned on a copper grid.

TEM micrographs were acquired using a transmission electron microscope (JEOL JEM-1400Plus) with (c g a s

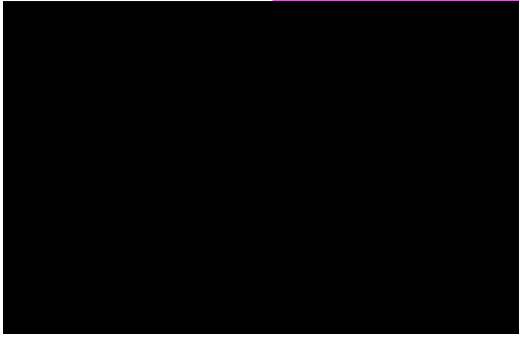


Figure 1: Nano-emulsion (O/W) of peppermint essential oil *M. piperita* (4%) obtained by (A) Before Sonication, (B) Sonication.



Figure 2: Density distribution diagram of *M. piperita* nano-emulsion sonicated for 45 min with oil and surfactant ratio of 1:4

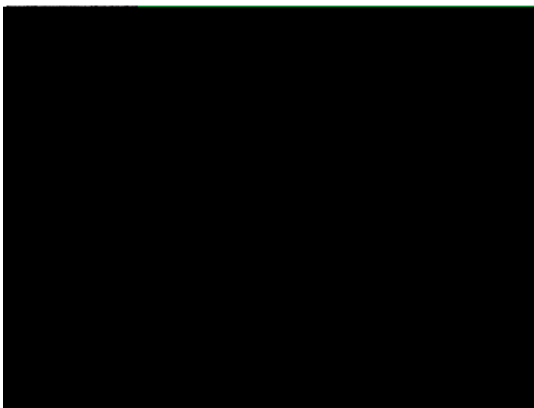


Figure 3: Round, spherical in shape, a good dispersion and narrow size distribution of peppermint oil *M. piperita* nano-emulsion at 4% by Transmission Electron Microscopy (TEM).

Contact toxicity bioassay using thin film residue

Insecticidal activity of *M. piperita* nano-emulsion and the free *M. piperita* EO were evaluated by thin film residue contact toxicity against *S. oryzae*. Results showed that both of them have toxic activity against *S. oryzae* at 24, 48 and 72 h of treatment. The lethal concentration (LC_{50}) accounted 0.181, 0.147 and 0.136 $\mu\text{l}/\text{cm}^2$ of free *M. piperita* EO against *S. oryzae* at 24, 48 and 72 h post exposure, respectively, as shown in Table 1. However, the lethal concentration (LC_{50}) of *M. piperita* nano-emulsion accounted 0.127, 48 h 0.106 and 0.095 $\mu\text{l}/\text{cm}^2$ at 24, 48 and 72 h of exposure, respectively, as shown in Table 2.

Time	LC_{50} $\mu\text{l}/\text{cm}^2$	Confidence Limits		Slope	χ^2	Toxicity Index (TI)
		Lower	Upper			
24 hr	0.181	0.159	0.199	3.861 ± 0.451	1.132	
48 hr	0.147	-	-	3.696 ± 0.565	12.065	
72 hr	0.136	0.109	0.155	4.063 ± 0.674	3.668	1

for the *M. piperita* nano-emulsion formulations compared with the bulk *M. piperita* oil (Figure 5).



Figure 4 of free *M. piperita* EO against *S. oryzae* adults 24, 48 and 72 hr.

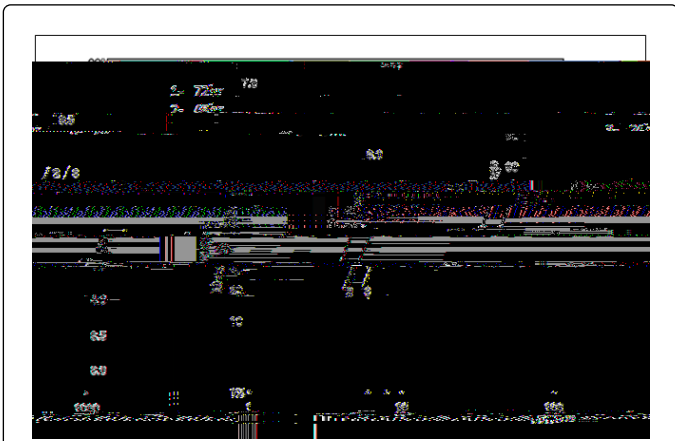


Figure 5 of the *M. piperita* Nano-emulsion against *S. oryzae* adults 24, 48 and 72 hr.

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- their nano-emulsions against *Callosobruchus maculatus* (F.). *J Stored Prod Res* 61: 9-16
29. Abbott WS (1925) A method of computing the LD_{50} of an insecticide. *J Econ Entomol* 18: 265-267.
 30. Qian C, Decker EA, Xiao H, McClements DJ (2012) Physical and chemical stability of β -carotene-enriched nano-emulsions of pH, ionic strength, temperature, and emulsifier type. *Food Chemistry* 132: 1221-1229.
 31. Ostertag F, Weiss J, McClements DJ (2012) Low-energy formation of edible nano-emulsions: factors affecting droplet size produced by emulsion phase inversion. *J Colloid Interface Sci* 388: 95-102.
 32. McClements DJ (2002) Colloidal basis of emulsion color. *Curr Opin Food Sci Technol* 7: 451-455.
 33. Pey CM, Maestro A, Solé I, González C, Solans C, et al. (2006) Optimization of nano-emulsions prepared by low-energy methods at constant temperature using a factorial design study. *Colloids Surf A Physicochem Eng Asp* 288: 144-150.
 34. Sh A, Abdelrazeik AB, Rakha OM (2015) Nano-em