



completely how pollutants interact in natural heterogeneous soil systems in real high-concentration cases. The top of this is to have a universal method to face all possible cases of this and similar kind and predict their dynamics and interactions. The work presented here is to set the first steps to solve and meet this ambitious and leading goal.

## Materials and Methods

### Sampling methods

In this section the main aspects on sampling during the whole process of this research in solid, liquid and solid-liquid phases are described.

#### Soils

##### Field scale

Seven random sandy soils were used to carry out different tests in this work. These were taken from different areas in central Asturias of Spain at random spots as follows (GPS coordinates, latitudes and longitudes respectively): a. 43° 35' 12.1812", -5° 56' 35.0412"; b. 43° 21' 18.396", -5° 52' 18.5586"; c. 43° 21' 14.832", -5° 52' 18.5298"; d. 43° 33' 4.572", -5° 37' 19.8402"; e. 43° 32' 55.8666", -5° 37' 25.6836"; f. 43° 21' 6.2166", -5° 52' 30.561"; g. 43° 21' 10.476", -5° 52' 26.0142". For the collection of each sample herbs and soil top layers were removed with a hand fork. Each sample was taken from field surface with a maximum depth of 20 cm with a hand trowel. The samples were collected in plastic bags and transported immediately to the laboratory

$$\ln q = \ln k + \frac{1}{n} \ln C$$

Equation 3 - 3

2° Get the equivalent linear equation from equation 3 - 3:

$$\ln q = \ln k + \frac{1}{n} \ln C$$

Equation 3 - 4.

where  $\ln q = y$ ,  $\ln C = x$ ,  $\ln k = a$  and  $\frac{1}{n} = b$

3° Do the graph from equation 3 - 4 and get the constant values of k and n by intersection values of y versus x for this equation, of the inverse of logarithm and fraction.

t Constant adsorption

Constant adsorption equation was used to test and show the

### Experimental tests

Experimental system was designed in different tests in order to determine the effect of temperature, pH, stirring speed and salts on bio-availability, environmental quality for microbial growth or phenol adsorption. These tests were carried out in slurry reactors with polluted sandy soils. Each test was conducted in triplicate and its results were represented in mean values.

Soil samples were placed in 250 mL Erlenmeyer flasks (reactors), in different solid/liquid (w/w) ratios, depending on the test. Initial phenol concentrations were prepared from appropriate dilution of a phenol standard solution (2000 mg/L). The same procedure was followed when nutrient salt solutions were considered.

Reactors were placed in an orbital incubator, New Brunswick G.25 model, protected from sunlight. To avoid interferences due to microbial degradation processes, sodium azide 0.02% (w/w) was used as antimicrobial agent. The tests were as follows:

**pH and salts:** Different salt solutions were used to control systems pH, their compositions are shown in Table 1. Each solution is described by letter A, B, C, D, E or F. Even though all soils were used for this test, its focus was on samples with a pH approximately lower or equal to seven (pH  $\leq 7$ ), in this case soils T4, T5 and T6 as it is shown in Table 1. Other experimental conditions were kept constant: temperature equal to 28°C, S/L equal to 0.2 and stirring speed equal to 150 rpm.

#### Slurry reactor

##### t Soil concentration

The effect of soil concentration on bio-availability and adsorption of phenol was tested at S/L interval from 0.01 to 0.82 (weight/weight relation). During this test other experimental conditions were kept

constant: pH equal to  $7 \pm 0.5$ , temperature equal to 28°C and solution E (Table 1). The soil sample for this test was T1.

**Stirring speed:** The effect of stirring speed on bio-availability and adsorption of phenol was tested at the interval  $150 \pm 50$  rpm (the circular motion had as a radius of 3 cm). During this test other experimental conditions were kept constant: pH equal to  $7 \pm 0.5$ , S/L equal to 0.2, temperature equal to 28°C and solution E (Table 1). Soil samples were T1, T2 and T3.

**Isotherms:** The same experimental conditions were used to find the different isotherm equations for soils T1, T2, T3, T4, T5, T6 and T7. Solution F, described in Table 1, was used to carry out this test with initial phenol concentrations between 30 to 300 mg/L. Other experimental conditions were kept constant: temperature equal to 28°C, stirring speed of 150 rpm and S/L relation equal to 0.2.

**Temperature:** System temperature was evaluated at two different intervals. The first interval corresponded to slight variations, it was around  $28 \pm 2^\circ\text{C}$  for soils T1, T2, T3, T4, T5, T6 and T7 (Table 2). Its specific experimental conditions were S/L equal to 0.2 and solution E (Table 1); the second one was conducted with greater variations, it was around  $27.5 \pm 22.5^\circ\text{C}$  for soil sample T1. Its specific experimental conditions were S/L equal to 0.1, solution F (Table 1) and different phenol initial concentrations in an interval from 27 to 134 mg/L. Other experimental conditions were kept constant in both tests: pH equal to  $7 \pm 0.5$  and stirring speed equal to 150 rpm.

**Salts:** The effect of salts (taking into account their nature and concentrations) on the phenol bio-availability and soil adsorption capacity was conducted by study of adsorption equilibrium on soil sample T1. Salts, such as sodium chloride (NaCl), magnesium sulphate ( $\text{MgSO}_4$ ), ferric chloride ( $\text{FeCl}_3$ ), potassium hydrogen phosphate ( $\text{K}_2\text{HPO}_4$ ), sodium carbonate ( $\text{NaHCO}_3$ ) chloride ammonium

Compounds	Liquid solution characteristics					
	A	B	C	D	E*	P*
Phenol (mgt)	300	300	300	300	300	+
Sodium azide (c..i. wt/wt)	0_02	0_02	0.02	0.02	0.02	0.02
NaHCO <sub>3</sub> (g L)	- Tc 17.299 0S12g L) g L) 17.20281H389.391IC420 81H 9.4510.003(0.02)9L)0.0LC Cm5 TcG61 m 03(0.01 366 02420 81H 9.FeC13					



samples had low concentration of organic matter (OM) as it is shown in Table 2, it was less than 0.7% in most of them. Only two soils, T1 and T7, had organic matter above this value, around 3.78 to 2.98% respectively.

These results showed that soils T2, T3, T4, T5 and T6 had very poor conditions for plant and microorganism growth according to the literature [36,37]. Also, in the same way, these samples with less than 0.7% of OM did not have very good encouraged granulation, tilth, porosity, bulk density, water in filtration and availability, plasticity, cohesion, adsorption capacity and nutrient source [38,39].

It has been reported that similar soils, like soils T1 and T7, had relative buffering capacity around its pH value for its OM concentration [36]. These were around the value of strongly alkaline, 9.16, to moderately alkaline, 8.01, respectively for these samples.

Additionally, it was not clear that OM was the main factor to determine the CEC value of all soil samples. Only for soils with high OM concentration, soils T1 and T7, this variable had a linear increment, where a high CEC value meant a high OM value. In other cases, specially with soils T2, T4 and T6, with intermediate CEC values, OM concentration did not have a clear relation with it. This was, probably, due to its characteristics, nature or other soil properties. Moreover, the effect of OM concentration over other soil variables in soil samples was also not evident at naked eye to its complex nature [38].

Electrical conductivity Soil samples had different electrical conductivities (EC) in this order: T4>T7>T3>T6>T2>T1>T5 as it is shown in Table 2. This was related to soil dissolved components because they carried electrical current in liquid phase under experimental conditions.

The relation of EC and pH of soil samples was not clear because soils with high conductivity had neutral alkaline pH values, like soils T4 and T7. Also samples with low EC had high and low pH values, like soils T1 and T5. These factors, maybe, were more related to soil nature and its composition [40].

Soils with high EC had high CEC values like soils T2, T3, T4, T6 and T7. It is important to highlight that it has been reported that soils with high EC have high bioavailability of their nutrients, cations and anions, as dissolved salts [41-45]. Nevertheless, in this sense, experimental EC values below 200  $\mu\text{S}/\text{cm}$  were a clear indication that there were not enough available nutrients in some soil samples and it could show low organism activity, especially for soils with low EC and low OM at the same time, like soils T2, T3, T4, T5 and T6.

Other important variable related with EC was the clay concentration. Soils with the highest EC had the highest clay%, like soils T3, T4 and T7. This is so because the clays were the support of cations that later were measured in dissolution with other anions by EC [46,47].

Volumetric density: The volumetric densities of soil samples were around 0.7 to 1.30  $\text{g}/\text{cm}^3$  as it is shown in Table 2. The interrelation of this variable with pH and CEC was not evident for all soil samples [42]. This factor was more related with sand and silt proportions and their packing arrangement [48]. Soil samples with the highest sand and silt percentage had approximately the highest volumetric density, that was the case of soils T5, T2 and T1. However, this was not a linear correlation in all soil samples due to soil morphology, especially for soils T6, T7, T4 and T3.

It has been reported that high soil volumetric densities are related indirectly with poor nutrient and environmental conditions to support

In this section the main previous considerations to have stable and reproducible conditions during the experimental tests are mentioned. This is structured in two main subsections: (1) pH and salts and (2) soil concentration and stirring speed. As it can be deduced by its names in these subsections are described the effect of pH, salts, soil-liquid relation and stirring speed on phenol soil adsorption equilibrium.

Given the situation of complex natural soil samples, as it has been shown in previous section, it was required to have a set of experimental conditions where a comparative and reproducible analysis could be done with independence of soil sample.

The object of this was to find out which pH, salts (type and concentration), solid-liquid relation and stirring speed had to be used to have reproducible and comparative results during the experimental tests.

test situations where. Tw T\* [(521.83 Td [( )4(e r)lin pni)-5(l s)-6(a)9(m)uenmni en and sducic9 0.0tetmi12(ue)-5(ir)1

The next step was to increase the bicarbonate salt concentration to 500 mg/L in systems as it is described in table 1, solution B. As it can be appreciated in Figure 2, letter D, this increment in all cases allowed increasing the pH and stabilized its value in the required pH interval of 6 to 9 after only 12 hours. Nevertheless, for the media of soil T6, the pH stabilization was not reached even after 72 hours under this condition. Despite this result, the mean pH value in all studied systems after this test was  $7.9 \pm 1.56$ . In Figure 2, letters A and B, it can be seen how the pH of media under natural conditions (represented by letter A) changed to the new pH values due to the addition of bicarbonate salt. (represented by letter B).

In order to have media pH of soil T6 around the required pH interval, it was required to increase the bicarbonate salt to 600 mg/L as it is shown in Table 1, solution C. As it can be seen in Figure 2, letter C, the increment bicarbonate salt to this concentration allowed reaching the required pH interval for all samples in less time, especially for soil T6. This pH value was around  $8.36 \pm 0.45$ . The pH equilibrium for T4 and T5 was reached after just two hours, ten hours less than the previous test. In case of media of soil T6, this pH equilibrium was reached after just 24 hours, 48 hours less than the previous test. An important factor to highlight in this point is that this salt concentration allowed sustaining the pH equilibrium without significant variation independent of soil nature. This last interesting conclusion was the reason for choosing 600 mg/L as an ideal bicarbonate concentration for subsequent tests.

When the effect of other salts in the system was evaluated (solution D, Table 1), like  $\text{FeCl}_3$ ,  $\text{K}_2\text{HPO}_4$ ,  $\text{NaHCO}_3$ ,  $\text{NH}_4\text{Cl}$ ,  $\text{K}_2\text{SO}_4$  and  $\text{CaCl}_2$  [60-



achieving two important things: similar operating condition for pH and salt concentrations in all cases independently of their nature and the possibility to compare all the results. This is with the idea to have robust conclusions in this work.

Soil concentration and stirring speed: Stirring factor determines the relation between soils, solvents, microorganisms, nutrients and pollutants in ex-situ soil bioremediation studies. The start point is to know how the operation conditions influence and change the soil adsorption equilibrium to avoid any misinterpretation of initial conditions in the global process [5].

#### t Soil concentration

Different soil concentrations were tested to analyze the influence of this factor on total phenol soil adsorption equilibrium in sandy soil samples.

There is a trend in soil ex-situ bioremediation to use indistinctly soil concentrations without any consideration of this factor [9,14,65] and in many cases it is still unclear how the optimization of this variable can improve the process goals. Even when this factor is considered in these processes the effect of this variable on soil adsorption properties are poorly described in real cases for sandy soils [30,66,67].

These tests were carried out by putting different solid/liquid (S/L, relation weight/weight) relations under the same experimental conditions to determine phenol adsorption kinetics and equilibrium point as it is shown in section 3.4.2.1. The results of these showed that it is possible to reach the adsorption equilibrium after 6 hours independently of the S/L relation in the system (the results are not shown).

Even when the equilibrium was reached almost at the same time in all tests, small differences were observed about phenol adsorption equilibrium for each S/L relation. Only for a small interval, from 0.01 to 0.11 S/L relations (0.01  $\leq$  S/L  $\leq$  0.11), was possible to have approximately ideal suspension conditions for particles, this being without significant interference between them for adsorption factors. Even for cases with S/L equal to 0.2 approximately (S/L  $\approx$  0.2) these differences were reasonably negligible for this heterogenic case. They were around 20 mg/L in mean terms. The same tendency has been reported by Fukui et al. [68] in tests with different S/L relations. This means that for any point under similar conditions, the equilibrium is constant independent of the S/L relation.

Also during these tests it was observed that soils formed conglomerate structures in the centre of flasks almost immediately after starting the tests (less than 1 hour). The high density of particles was the main reason, even when the stirring speed and turbulence of the media were high, where the Reynolds number was equal to 13860 (NRe=13860) [21]. This effect was more significant when the soil-liquid relation was over 0.2 (S/L>0.2). Likewise, this soil conglomeration provided different experimental conditions during the adsorption process in the same reactor; there were particles in suspension, semi-

20]. More studies related to the adsorption equilibrium of contaminants in natural soil systems have been done for years. Most of them are specialized in adsorption equilibria where the pollutants and soils are related in ideal conditions with low pollutant concentrations and high soil adsorption capacities, such as activated carbon, polymeric resins or pure minerals [20,75-80]. However, heterogeneous soil adsorption processes with high loadings of pollutants have been poorly evaluated due to its complexity [81-83].

The object of these tests was to find an equation that it could be used to describe phenol soil adsorption isotherms independently of sandy soil nature in an ex-situ bioremediation process before microbial activity.

In order to carry out these tests, different soil samples were settled in different reactors under the same experimental conditions. This allowed us to do a real comparative analysis of the efficiency of tested equations. The specific details of this are described in section 3.4.3. of methodology section. In order to fit the equations to experimental data a graphical representation method was used as it is shown in section 3.3.1.

## Equilibrium

In ex-situ bioremediation processes, it is necessary to know how the pollutants are distributed around phases to find the best experimental conditions to achieve process goals. Without this consideration, it is not possible to predict the availability of pollutants in the liquid phase and know if the microorganisms can degrade them easily under a set of experimental conditions.

This section of equilibrium is structured in two subsections. They are related with results of isotherm equations and soil properties related with phenol adsorption equilibrium.

The key factor, before microbial activity in a bioremediation process, is to predict the adsorption behaviour and understand its properties, in the equilibrium solvent-solute-adsorbent, to implement all required actions to quickly recover the polluted soil. There are a lot of studies about how to predict adsorption behaviour with different isotherm equations and about which soil factors are more important in this process in ideal solid-liquid conditions. However, natural soil bioremediation cases with high pollutant concentrations are poorly evaluated.

The object of these tests was to find a universal isotherm equation to predict the adsorption behaviour of high phenol concentrations in natural sandy soils and find which soil properties can influence this adsorption equilibrium under real bioremediation conditions before microbial activity.

Isotherms: Equations used to describe the adsorption isotherms in solid-liquid systems are derived from the models developed for solid-gas systems [17]. Despite this, excellent results have been obtained to represent real cases of this kind in low concentration or in conditions where the nature of the adsorbent is homogeneous. The challenge here is to find equivalent equations for heterogeneous conditions in natural environments like those that are easily to find in soil bioremediation processes.

In this section, results of equation validations for phenol soil adsorption isotherms during experimental phase are shown.

Even adsorption isotherm equations that come from solid-gas systems, as it was mentioned above, there are many of them that are especially improved for ion exchange studies in solid-liquid systems [18-

The correlation order for phenol adsorption isotherms was different for each soil sample and each tested equation. The order of accuracy between the experimental data and calculated data was bigger in the case of the Freundlich equation in relation to others. The fit of the experimental data to tested equations was as follows, in the order of accuracy from the highest to the lowest: Freundlich (0.95) > constant separation factor ( $\neq 0.91$ ) > linear adsorption model ( $\neq 0.21$ ).

The small variation between Freundlich and separation constant isotherm, represented by correlation factor of 0.95 and 0.91 respectively, showed that both equations can be used to represent the

factors in unced phenol adsorption equilibrium for some soils. For that reason additional analyses were needed to clarify this issue. is analysis is presented in the following section.

Soil properties:Standard protocols for ex-situ soil bioremediation processes are needed in order to achieve the process goals independently of soil nature and environmental conditions. As it was mentioned, the soil properties and their control can be key factor to enhance availability of pollutants in medium if it is analyzed correctly in soil adsorption process.

In this section, a comparative analysis of soil properties and how these in uenced the phenol soil adsorption process for studied soil samples is described.

In ex-situ soil bioremediation processes the pollutant distribution in the soil determines the guidelines to follow in the cleaning of these systems. Most studies which are related to this issue are focused on groundwater contamination, where contamination levels are generally low [10,80,82,83,86]. In these and other similar studies, the highest correlation coe cients (r) of adsorption processes generally correspond to the tests conducted in systems with a high concentration of organic matter. Other factors prevailing in this connection is the percentage of clay and cation exchange capacity [25,75,87-90]. However, it is not completely clear how these factors in uence adsorption conditions with heterogeneous natural soils in high phenol concentrations.

e object of this analysis was to nd which soil properties could in uence the adsorption equilibrium under real bioremediation conditions before microbial activity.

e experimental data used to carry out this analysis were taken from the test described in the previous section (5.3.1) All details of experimental conditions and related information are described there.

e experimental results showltio thidwa 5.247 13d4(o)(en)19(t)11(u)-5(ld )JTJue2fith hhe highest roperpion of organic atter5OM),c elywy tpu.

)16(n t)-3(a)1(s)-8(e)of ooil pOM,7 13d4it is nhowlanin tTlet 2, rom tahhe -5(o)11(r)13(e)iec-3(a)-5(l a)-9(e)12(ri)19(t) ff oiovw73(.,Ed.9381 523.7578 Tm (2T1, T4, T6a)9(n)4(d cT7n)4(h)4(o)16(u)-5(ld )n a34-7(v)8(erhn a34dh)-6(h)4(e hig)-7(h)4(es)5(t) ehenol corhese fd.9381 523.7578 Tm (2)4(h)4(o)16(u)-5(ld )n a34-7(v)8(erh)-9(e)-5(den:o)-8(o)12(i)-5(l pT1>)-8(o)12(i)-5(l pT7>)-8(o)

ene mn3(o)16(ut)howlanin t8(e)-ns cwork it ias tn3(o)16(ut)etslet d o-1(u)-2(t i)-8(v)83(i)dtnd cedioldtteaieneyw8(e)-85(enfd.9381 523

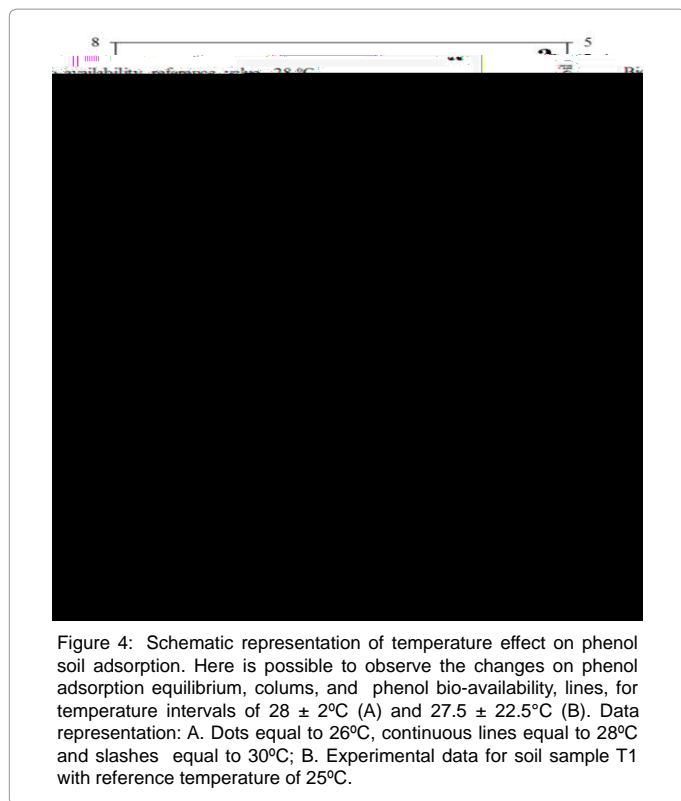


Figure 4: Schematic representation of temperature effect on phenol soil adsorption. Here is possible to observe the changes on phenol adsorption equilibrium, columns, and phenol bio-availability, lines, for temperature intervals of  $28 \pm 2^\circ\text{C}$  (A) and  $27.5 \pm 22.5^\circ\text{C}$  (B). Data representation: A. Dots equal to  $26^\circ\text{C}$ , continuous lines equal to  $28^\circ\text{C}$  and slashes equal to  $30^\circ\text{C}$ ; B. Experimental data for soil sample T1 with reference temperature of  $25^\circ\text{C}$ .

Temperature: The effect of temperature on soil adsorption capacities of pure compounds with high content of organic matter has been widely described in literature. Presently, due to the great amount of polluted soils around the world, it is the time to know its effects on real polluted soil environments.

In this section the results of the effect of temperature on adsorption and bio-availability of phenol in sandy soil samples are presented.

In many phenol bioremediation processes it is common to see changes in temperature that enhance microbial activity. However, the effect of this variable on soil adsorption properties is unconsidered during this process in many cases [98,99]. Little is known about the effect of temperature on the adsorption equilibrium of phenol in natural sandy soils under high phenol concentration cases.

The object of these tests was to find how temperature could influence the adsorption and bio-availability of phenol during a process of ex-situ soil bioremediation in sandy soils before microbial activity.

Two different intervals of temperature were tested in this sense, one around  $28 \pm 2^\circ\text{C}$  and another one around  $27.5 \pm 22.5^\circ\text{C}$ . The other experimental conditions were kept constant and are described in section 4.4.4. The theoretical isotherms were represented by Freundlich equation.

The experimental data in the first set of tests showed, represented in Figure 4A, that small changes in system temperature, around  $28 \pm 2^\circ\text{C}$ , caused slight differences in soil adsorption capacity and bio-availability of phenol for each soil sample. In the case of soil adsorption capacities (columns), these data showed that for four soil samples, T1, T2, T3 and T5, an increment in system temperature meant an increment in phenol adsorption with respect to the equilibrium. However, the opposite behaviour was observed for soils T6 and T7, where an increment system temperature meant a decrement of phenol adsorption with

respect to the equilibrium. For soil sample T4, the change of system temperature in this interval did not affect significantly its adsorption capacity, its variation has been less than 5%; in case of phenol relative bio-availability with respect to the phenol equilibrium concentration (lines), it was noted that small differences between equilibrium conditions in that range of temperature were around  $\pm 2.5\%$  for each case in mean terms, as it can be seen in this figure. It meant that almost all equilibrium conditions were the same in that interval of temperature. The average standard deviation (SD) of concentration for each soil in this range of temperature was:  $T1SD = \pm 4.19$ ;  $T2SD = \pm 2.45$ ;  $T3SD = \pm 2.49$ ;  $T4SD = \pm 0.94$ ;  $T5SD = \pm 2.05$ ;  $T6SD = \pm 1.64$ ;  $T7SD = \pm 5.35$ . And for all soil samples together under these conditions it was  $\pm 2.72$  mg/L at equilibrium concentration. These small differences did not allow developing the real nature of this phenomenon. For this reason it was necessary to carry out more specific tests for this work.

The second set of tests about the effect of temperature on phenol adsorption capacity in the interval of  $27.5 \pm 22.5^\circ\text{C}$  showed, as it can be seen in Figure 4B, that an increment system temperature meant a decrement of phenol adsorption with respect to the equilibrium (columns). Its consequence was that there were more phenol molecules available in liquid phase for soil sample T1 (line). Between temperatures  $5$  and  $1^\circ\text{C}$  it was noted that an increment in the system temperature meant an increment in soil phenol adsorption with respect to the equilibrium concentration. The reason of this behaviour, that it was the opposite of previous one, was probably due to the interrelation of phenol with some soil components whose concentration in liquid phase it had reduced [100,101]. Additionally, it was observed that reproducibility of results in this interval has had the highest standard deviation and this could be the reason of this behaviour even when triplicate analyses were done. For this reason an extra test on phenol adsorption was carried out.

When other experimental conditions to get the phenol adsorption isotherms for this soil in the same interval of temperatures were tested, more evident results were obtained, and they are represented in Figure 5. When the system temperature decreased, from  $50$  to  $15^\circ\text{C}$ , phenol soil adsorption equilibrium was displaced. This change in the system

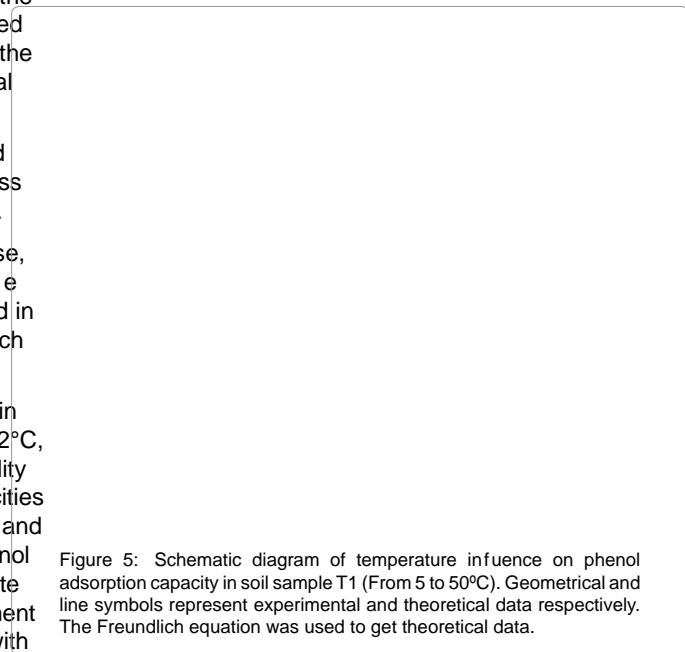


Figure 5: Schematic diagram of temperature influence on phenol adsorption capacity in soil sample T1 (From 5 to  $50^\circ\text{C}$ ). Geometrical and line symbols represent experimental and theoretical data respectively. The Freundlich equation was used to get theoretical data.

temperature increased the retention of phenolic molecules in the soil adsorption and reduce its bio-availability. It is still required to know if the reason of this could be that at high temperature there was an "the remedy is worse than the disease" in this sense. In this section the results of salts effect on phenol soil adsorption equilibrium in an ex-situ bioremediation process before microbial activity are presented.

Also these data and their analysis, as it is shown in Figure 5, allowed finding out the complex behaviour of phenol soil adsorption at 5°C. Organic soluble or gaseous compounds inert to the environment and the reason for this could be, as it was mentioned before, the reaction innocuous to human health. During this process, the microorganisms of phenol with some salts in the soil structure. Nevertheless, its natural capability of consuming these contaminants quickly increase their number. Critical micro and macro nutrients, or bio-stimulants, are and composition was undetermined. Other more specific tests have to be done in order to understand this phenomenon. required in this process, like carbon, nitrogen, phosphorus, potassium, sodium. Without these nutrients, umail 411

The results of these tests showed that system temperature could enhance the availability of phenol in the liquid phase and reduce the phenol concentration in soil if there is an increment in temperature. Further, these results showed that the characteristics of this equilibrium are determined by the interrelation of phenol-water-soil. Without this information, it is impossible to find out this behaviour without empirical tests in each case. Other additional tests are required to find out what happens with soil adsorption properties in systems with high charge of phenol and heterogeneous soils at low temperatures.

Salts: Nutrient salt applications are common practice in soil bioremediation processes because these can enhance the microbial activity and reduce the recovery time. However, in all cases it is not completely clear if these substances can increase the pollutant soil

After reaching the phenol adsorption equilibrium, the isotherms were determined by Freundlich equation.

These tests and results showed that phenol soil adsorption process in real environments is complex and difficult to predict. The salts, by kind and amount, increased, increased-decreased or decreased the phenol soil adsorption capacity. Also, these results have shown a competitive multilayer adsorption tendency. These results, represented by its isotherms, are shown in Figure 6.

These results showed that an increment of  $K_2HPO_4$  salt around 12 mg/L did not alter significantly the phenol adsorption equilibrium. However, concentrations of this salt, above this value (>12 mg/L), increased considerably the phenol retention in soil matrix and reduced

#### Acknowledgment

Gularte F. thanks the economic support of all his family, especially to his grandparents Guadalupe and Francisco, Lorena (mother), brother (Hector), Ronald, and the Spanish Agency of International Cooperation (AECI). Also, this author thanks all his supervisors, colleagues and support staff for their support in this amazing learning experience at the University of Oviedo. Gularte F gives special thanks to Mr Fernando Gonzalez for the web links that he provided for this work. Gularte also thanks all people that he found in his way during this work because in certain ways they indirectly contributed to this work.

#### References

1.

---



89. Malusis M,