

Keywords: Phenol; Biodegradation; *Pseudomonas aeruginosa*; Plackett-Burman Design; Central composite design

Introduction

Phenol is one of the most common toxic environmental pollutants, arising during the processing of resins, plastics, dyes, varnishes, pharmaceuticals and pesticides from several industrial processes and several chemical industries [1]. Phenol concentrations are toxic to fish if it is higher than 2 mg/L and concentrations between 10 and 100 mg/L would result in death of many aquatic lives within 4 days [2]. The Environmental Protection Agency (EPA) stated that less than 1.0 ppb of phenols in surface waters is good for health. The World Health Organization (WHO) has set a limit level of 1 mg/L to regulate the phenol concentration in drinking waters [3].

Therefore, development of new methods has generated significant interest for phenol removal from industrial wastewater [4]. Chemical or physical methods for phenolic waste treatment were used conventionally such as chemical oxidation, solvent extraction and adsorption, but these processes are facing secondary treatment problems. Apart of these methods, biological treatment i.e. Biodegradation is good alternative. Biodegradation is versatile, inexpensive and can potentially turn a toxic material into harmless products [5].

The use of pure cultures microorganisms, especially for the efficient metabolism of the contaminant, is advisable as a better alternative compare to mixed culture [6-9]. In general, the one-variable at a 9(en)8(sina(l)(i TJ 0.025 Tw T* [(o)12(f va)9(r)-6(i)-3(a)8(b)7(les)

was applied to evaluate nine factors (including two dummy variables) suggested by [15]. The medium was sterilized in two parts to avoid precipitation of ferrous salts during autoclaving. These were denoted as Part A and Part B with the following compositions:

Part A K_2HPO_4 , KH_2PO_4 , $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$

Part B $(\text{NH}_4)_2\text{SO}_4$, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, MnSO_4 , H_3BO_3 , NaCl

After autoclaving, the contents were cooled and then mixed together and kept as stock solution from which known quantities were taken for the culture.

The pH of the medium thus obtained was 7.0 ± 0.1 .

Acclimatization

The primary culture was transferred to the two loops for acclimatization.

The concentration of phenol degraded in the solution was determined by a UV-vis spectrophotometer using 450 nm wavelength.

Plackett-Burman design

Plackett-Burman design, an efficient way to identify the important factors, was used for the optimization of the process.

levels below 95%, were considered insignificant and were not included in CCD experiments. The Pareto chart shows the significant variables in Figure 1.

The model equation for phenol degradation (Y) could be written as:

$$Y = 82.53 - 1.14 A - 2.43 B - 11.53 C + 0.56 D + 25.75 E + 61.27 F - 13.72 G - 6.80 H - 0.33 J$$

Optimization by response surface methodology and regression analysis

CCD was employed to study the interactions between the significant

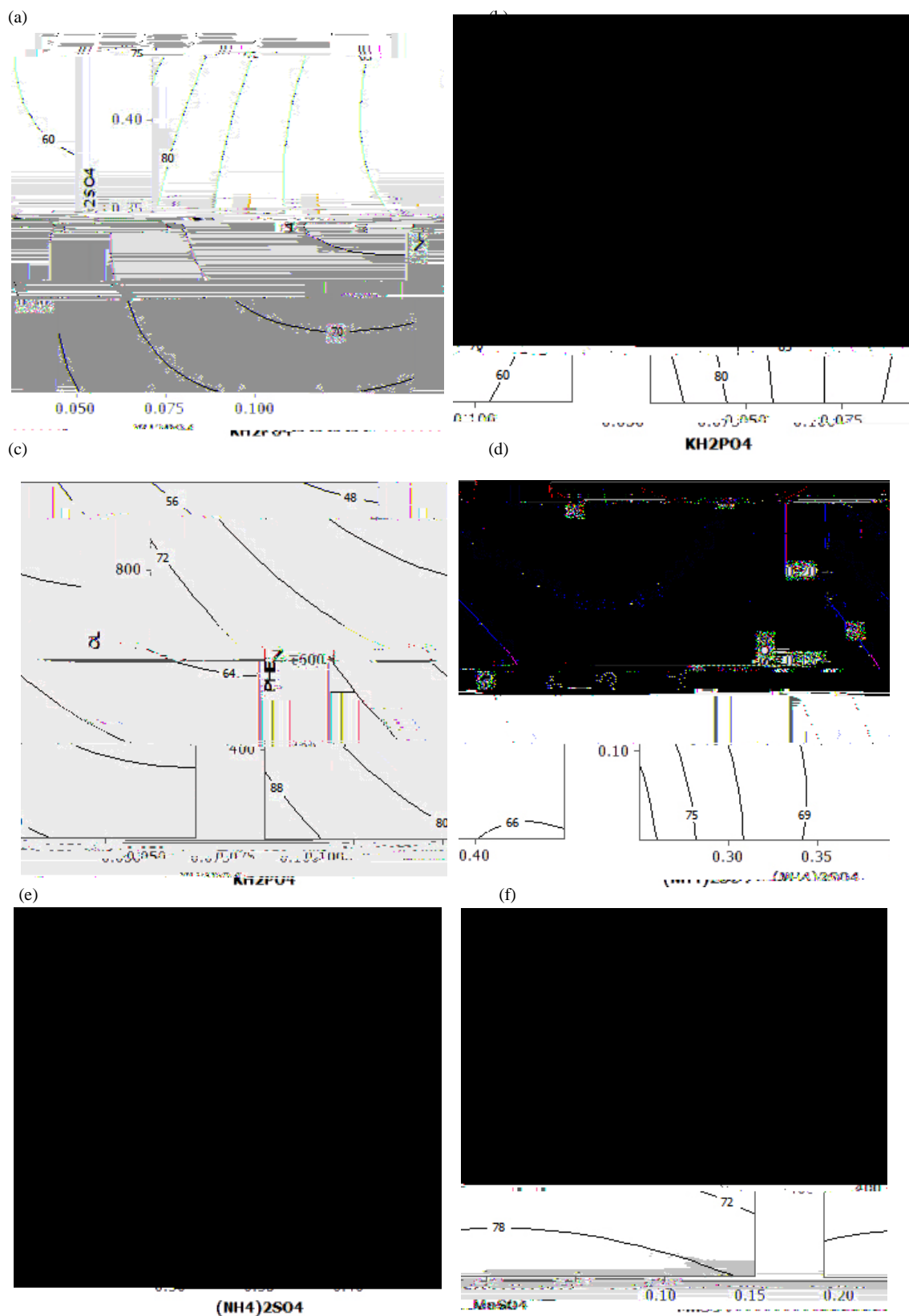


Figure 2: Contour plots showing the interaction effect between pair of variables (a) KH₂PO₄ and (NH₄)₂SO₄, (b) KH₂PO₄ and MnSO₄, (c) KH₂PO₄ and phenol, (d) (NH₄)₂SO₄ and MnSO₄, (e) (NH₄)₂SO₄ and phenol, (f) MnSO₄ and phenol on phenol degradation (%) by keeping the other two variables constant at middle level.

of phenol. Four medium variables out of nine were identified to be significant on phenol degradation by PB experiment. The results obtained from RSM were clearly explained that optimum values of significant variables had a significant effect and promotes an increase in percentage of phenol degradation. The optimization results shows that the maximum phenol degradation of 81.62 (%) was obtained when KH_2PO_4 , $(\text{NH}_4)_2\text{SO}_4$, MnSO_4 and phenol were kept on optimized concentrations 0.05 g/L, 0.35 g/L, 0.15 g/L and 500 mg/L respectively.