

Keywords : RNA; RNAlater; TRIzol; RNase; Allprotect; PAXgene

Introduction

Gene expression analysis provides information at which a particular gene pattern may be expressed by cellular responses. Gene expression is evaluated by means of reverse transcription polymerase chain reaction (RT-PCR), quantitative real-time RT-PCR (qRT-PCR) as well as multiplex gene expression quantification arrays and, data analysis depends on the RNA integrity and stability isolated from cells [1]. RNA quality is influenced by warm and cold ischemic durations, cellular stress responses, tissue processing protocols and storage conditions [2,3]. Isolation of highly pure intact RNA is vital for successful quantification of gene expression [1,4] so that RNA preservation is an essential subject during handling processes [5]. Since RNA degradation takes place by cytoplasmic RNase [4] thus, RNase inhibition is a main approach in RNA extraction and subsequently gene expression analysis [1,4].

Conclusion

Abstract

RNA degradation correlates with the length of delay in sample fixation or preservation. Most pathology departments still rely on tissue preservation with formalin fixation followed by paraffin embedding step [3]. Although it is suitable for morphologic assessment, it is unable to keep intact RNA [5].

In fact, formalin fixative keeps tissue structure via cross-linking of cellular proteins. In addition, there also happen cross-links between proteins and nucleic acids. Mono-methylol groups (-CH₂OH) added to all four bases of RNA create methylene bridges between two amino groups. Although it makes formalin an ideal fixative hardening agent for histopathological analysis, cross-linking of proteins and nucleic acids causes intact mRNA extraction from formalin-fixed tissues to be difficult [3,4,6,7]. RNA extracted from formalin-fixed paraffin-embedded (FFPE) tissues is chemically modified and dramatically degraded. It is often present in fragments less than 300 bases in length [4,5,8,9] and may produce reliable data if selected genomic sections be heavily amplified using specific primers and short amplicons (<300 bps) [4,10]. In addition, RNA chemical modifications in FFPE tissues can be reversed with proteinase K digestion and preheating in guanidinium or TE (10 mM Tris-HCl, pH 7.0, 1 mM EDTA) or citrate (pH 4.0) buffers making it more amenable to RT-PCR yet, the major drawback of formalin fixation (i.e. fragmentation) remains uncorrected [4,9,10].

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